

# Agro-Industry Waste Fed Microbial Electrolysis Cell for Biohydrogen Production

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## Abstract-

The microbial electrolysis cell is gaining advantage over the other biological hydrogen production techniques as it requires less energy for hydrogen generation as compared to the water electrolysis process. The present study aims to assess the aptness of Agro-Industry Waste (AIW) fed membrane-less single chambered Microbial Electrolysis Cell (SC-MEC) for the biohydrogen production in batch mode under applied voltage of 1 V at  $30 \pm 2$  °C (Fig.1). The performance of the reactor was assessed through volume of hydrogen per gram of COD removed, coulombic efficiency, cathodic hydrogen recovery and COD removal efficiency. The highest COD removal of 71% was reported with coulombic efficiency of around 45%. These results demonstrated an energy-efficient approach for biohydrogen production from AIW coupled with waste mitigation

**Keywords:** Electro-hydrogenesis, Microbial Electrolysis Cell, Electrode Modification, Over-potential Reduction, Hydrogen Evolution Reaction

## 1. Introduction

The energy crisis along with environmental pollution e.g. greenhouse gas emissions etc. are among the major global challenges [1–3]. Fossil fuels are the major fuels sustaining the current energy needs. The world's focus towards sustainability to counter fuel scarcity and soaring prices, thus thrust for alternative renewable energy carrier is getting intensified[4,5]. The biological H<sub>2</sub> production methods are based on waste to energy route which gives this technology an edge. They are cheaper and environment friendly [6]. Hydrogen production methods vary from water electrolysis, thermo-chemical methods, biological methods etc. [7]. Microbial electro-synthesis systems (MES) are an efficient device for biofuel production [8,9]. The biological routes of hydrogen production are promising as they deliver hydrogen from waste as well as manages the waste and pollution. Hence, the scientific community is penetrating renewable energy resources to answer the energy scarcity through the waste energy route. This has led to an integration of solid waste management and biofuel production. Among the potential biofuels studied, Bio-hydrogen is considered as one of the most attractive alternatives owing to the properties of the hydrogen as fuel [10–14]. The combustion of hydrogen

produces only heat and water without any greenhouse emissions which makes it a potential fuel. The evolution of Microbial Electrolysis Cell (MEC) in recent times has gained momentum for H<sub>2</sub> production from inexpensive organic materials such as waste food materials, thermal and chemical hydrolysate, sludge, industrial effluents, landfill leachate etc. [15]. MECs are based on electro-hydrogenesis process of biodegradable materials [14]. They are anaerobic systems, can be either single chambered or double chambered. The exo-electrogenic bacteria at anode oxidize organic matter and transfer electrons extracellularly to cathode via external circuit, while proton travel through proton exchange membrane or directly to cathode. The catalyst if present, at cathode catalyzes the formation of H<sub>2</sub> from electrons and protons. Theoretically, MECs require only external voltage of around 0.11 V to drive the H<sub>2</sub> production from acetate [1,16].

The current study focuses on the utilization of Agro-Industry Waste (AIW) in batch mode of membrane less single chambered Microbial Electrolysis Cell (SC-MEC) for the biohydrogen production at an applied voltage of 1 V at  $30 \pm 2$  °C (Fig.1).

## 2. Experiments

### 2.1. Reactor design and construction

Single chambered MEC was constructed with acrylic sheet in a cube shape with side = 12 cm with a working volume of 700 mL. The carbon cloth electrodes of surface area (approx. 10 cm<sup>2</sup>) were kept at an effective distance of 6 cm to reduce the overpotential in MEC. A 100 Ω resistor was connected in series with the electrodes by copper wires.

### 2.2. Reactor Inoculation and operation

The electrode enrichment for bioanode was achieved with a pure culture of *Shewanella putrefaciens* on heat treated carbon cloth and then switched to fully anaerobic SC-MEC. The MEC was inoculated with pure culture digestate from the Microbial fuel cell fed AIW. After acclimatization AIW was fed to the MEC and purged with N<sub>2</sub> for 15 min before and after of feeding. The bio-film was

achieved on anode from MFC kept in MEC mode at 1 V for 72 h [2]. The phosphate buffer was used to maintain the pH.

### 2.3. Reactor performance parameters

The performance of the reactor was assessed through volume of hydrogen per gram of COD removed, coulombic efficiency, cathodic hydrogen recovery and COD removal efficiency. The highest COD removal of 71% was reported with coulombic efficiency of around 45%. These results demonstrated an energy-efficient approach for biohydrogen production from AIW coupled with waste mitigation. This lab scale study of 500 mL of SC-MEC resulted in the promising approach of hydrogen production

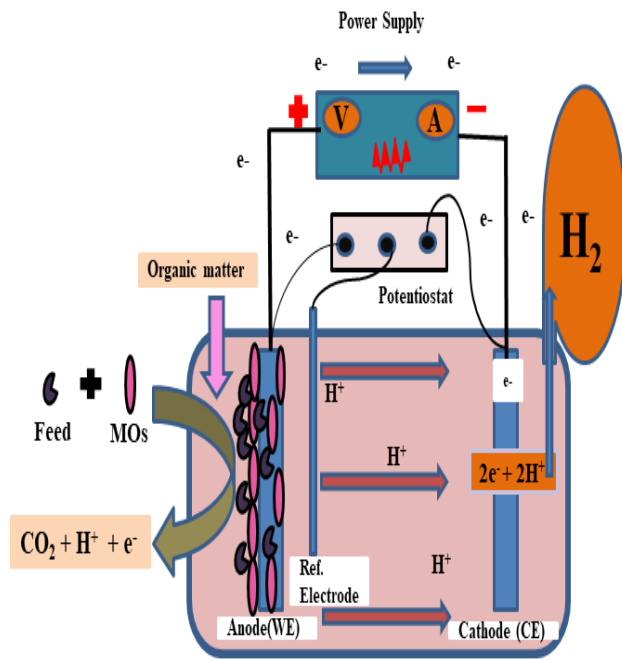


Figure 1. Schematic representation of MEC

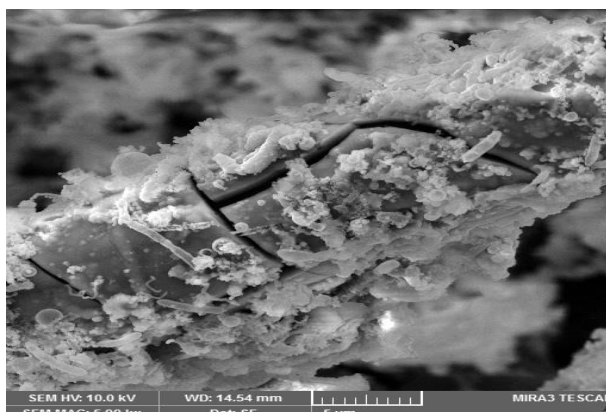


Figure 2. SEM image of bio-anode

and can be escalate to subsequent pilot scale studies. The FE-SEM image of bioanode had shown a biofilm cover over the carbon cloth (Fig.2).

### 2.4. Gas measurement, storage and analysis

The produced bio-hydrogen was measured by the well-established water displacement method. The gas was stored in a graduated measuring cylinder. After the batch cycle, produced gas was analysed by the Gas Chromatography by using a gas tight syringe and argon as the carrier gas in TCD mode.

## 3. Results and Discussion

### 3.1. MEC performance and Bio-film formation

After the acclimatization at 0.5 V, MEC fed with AIW was given 1 V in a batch cycle. The maximum current of 15 mA and corresponding current density of 5.5 A/ m<sup>2</sup> were achieved. The total volume of gas was recorded as 1890 mL with 64 % of H<sub>2</sub> content (by Gas chromatography). The bio-anode samples were scanned under Scanning Electron Microscope. The biofilm formation on the electrode surface can be seen in the image (Fig. 2).

### 3.2. COD removal and Substrate degradation

The organic matter of substrate reportedly degraded by microorganisms and reflected in terms of the COD removal, Total Suspended Solid (TSS), Volatile Suspended Solids (VSS) reductions (Fig.4) and eventual Hydrogen production. The initial phase resulted in lower COD removal but with the passage of time COD removal efficiency of the MEC increased continuously and at the end of the 21st day cycle, COD removal of 70.8 % was achieved.

### 3.3. H<sub>2</sub> production rate

The production rate for hydrogen was investigated to evaluate the performance of the MEC. A total of 1890 mL of gas was collected at the end of the batch cycle. The hydrogen production rate continuously increased as indicated by the COD removal percentage. The coulombic efficiency (CE) and the cathodic gas recovery (R<sub>c</sub>) are two parameters to evaluate the MEC performance along with the production rate [13,17].

Total amount of Hydrogen (V<sub>h</sub>) in total gas is calculated based on Eq. 1.

$$V_h = (H_s + V_t)G_f \quad (1)$$

Where –

V<sub>h</sub> - volume of Hydrogen in total gas

H<sub>s</sub> – headspace volume in mL

V<sub>t</sub> – total volume of gas in mL

G<sub>f</sub> – fraction of Hydrogen in gas measured by GC

The expected gas production (V<sub>expt</sub>) from the complex substrate is given by Eq. 2

$$V_{expt} = C_t * \frac{V_m}{2F} \quad (2)$$

where,

Ct – charge over the given time in Coulomb

V<sub>m</sub> – volume of one mole of gas in mL

F – Faraday Constant

The Cathodic hydrogen recovery (R<sub>c</sub>) is the measure of the conversion of electrons to hydrogen (Eq. 3). It is the ratio of V<sub>h</sub> to V<sub>expt</sub>. The R<sub>c</sub> is used to calculate the coulombic efficiency (CE) in Eq. 4. It is the ratio of current measured over a time to the theoretical current based on COD.

$$R_c = \frac{V_h}{V_{expt}} \quad (3)$$

$$CE = \frac{\eta_{ce}}{\eta_{th}} \quad (4)$$

Based on the results obtained from gas chromatograph, the amount of hydrogen in 1890 mL of gas produced and 200 mL of headspace, was estimated as 1690 mL. The results obtained were in accordance and comparable to other studies on waste as shown in Table. 1.

| Feed | E <sub>a</sub><br>p | CE<br>(%) | R <sub>c</sub><br>(%) | Q<br>(m <sup>3</sup> /m <sup>3</sup> /d<br>ay) | COD<br>remo<br>val % | Referen<br>ces |
|------|---------------------|-----------|-----------------------|--|----------------------|----------------|
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#### References

- [1] Kadier A, Jain P, Lai B, Kalil MS, Kondaveeti S, Alabbosh KFS, et al. Biorefinery perspectives of microbial electrolysis cells (MECs) for hydrogen and valuable chemicals production through wastewater treatment. *Biofuel Res J* 2020;7:1128–42. <https://doi.org/10.18331/BRJ2020.7.1.5>.
- [2] Gautam R, Nayak JK, Talapatra KN, Amit, Ghosh UK. Assessment of different organic substrates for Bio-Electricity and Bio-Hydrogen generation in an Integrated Bio-Electrochemical System. *Mater Today Proc* 2021;6–10. <https://doi.org/10.1016/j.matpr.2021.06.223>.
- [3] Kumar Nayak J, Gautam R, Uttam ·, Ghosh K. Bioremediation potential of bacterial consortium on different wastewaters for electricity and biomass feedstock generation. *Biomass Convers Biorefinery* 2022;1:1–14. <https://doi.org/10.1007/S13399-022-02992-2>.
- [4] Nayak JK, Amit, Ghosh UK. An innovative mixotrophic approach of distillery spent wash with sewage wastewater for biodegradation and bioelectricity generation using microbial fuel cell. *J Water Process Eng* 2018;23:306–13. <https://doi.org/10.1016/j.jwpe.2018.04.003>.
- [5] Amit, Kumar Ghosh U. Utilization of kinnow peel extract with different wastewaters for cultivation of microalgae for potential biodiesel production. *J Environ Chem Eng* 2019;7:103135. <https://doi.org/10.1016/j.jece.2019.103135>.
- [6] Hassan M, Fernandez AS, San Martin I, Xie B, Moran A. Hydrogen evolution in microbial electrolysis cells treating landfill leachate: Dynamics of anodic biofilm. *Int J Hydrogen Energy* 2018;43:13051–63. <https://doi.org/10.1016/j.ijhydene.2018.05.055>.
- [7] Jabbari B, Jalilnejad E, Ghasemzadeh K, Iulianelli A. Recent progresses in application of membrane

| Agro Industry Waste | 1   | 45      | 54.6     | 0.2          | 71        | Present study |
|---------------------|-----|---------|----------|--------------|-----------|---------------|
| Leachate            | 1   | 12-41   | 66-95    | 0.04-0.06    | 65-73     | [6]           |
| Acetate             | 0.8 | 8-42    | 65-93.8  | 0.034-0.237  | 86.6-97.5 | [18]          |
| Sludge              | 0.6 | NA      | NA       | 4.6 mg/g VSS | 17-53     | [16]          |
| Acetate             | 1   | 22.80 % | 101.4 0% | 0.3          | NA        | [20]          |
| Glycerol            | 0.8 | 35      | 4        | 0.021        | 100       | [22]          |
| Milk                | 0.8 | 52      | 13       | 0.086        | 73.5      | [22]          |

#### 4. Conclusion

The present study on the SC-MEC fed with agro industry waste resulted into 71 % of COD removal and 45% of coulombic efficiency. The results of this study has been compared with the other similar studies and has shown in table 1. The hydrogen production rate and current density of 0.2 m<sup>3</sup>/m<sup>3</sup>/day and 5.5 A/m<sup>2</sup> could be further increased by efficient extracellular electron transfer achieved through electrode modifications. This study indicated that AIW resulted into comparable or even higher coulombic efficiencies and COD removal in comparison to the most studied feed acetate. Also, enhanced rate of hydrogen production was achieved.

- bioreactors in production of biohydrogen. *Membranes (Basel)* 2019;9:1–30. <https://doi.org/10.3390/membranes9080100>.
- [8] Nelabhotla ABT, Khoshbakhtian M, Chopra N, Dinamarca C. Effect of Hydraulic Retention Time on MES Operation for Biomethane Production. *Front Energy Res* 2020;8:1–6. <https://doi.org/10.3389/fenrg.2020.00087>.
- [9] Kumar G, Bakonyi P, Zhen G, Sivagurunathan P, Koók L, Kim SH, et al. Microbial electrochemical systems for sustainable biohydrogen production: Surveying the experiences from a start-up viewpoint. *Renew Sustain Energy Rev* 2017;70:589–97. <https://doi.org/10.1016/j.rser.2016.11.107>.
- [10] Mogili NV, Murugesan N, Ayothiraman S, Gautam R, Deshavath NN, Reddy Erva R. Biohydrogen production from wastewater and organic solid wastes. *Waste-to-Energy Approaches Toward Zero Waste* 2022;1:65–95. <https://doi.org/10.1016/B978-0-323-85387-3.00009-4>.
- [11] Gautam R, Nayak JK, Daverey A, Ghosh UK. Emerging sustainable opportunities for waste to bioenergy: an overview. *Waste-to-Energy Approaches Toward Zero Waste* 2022;1–55. <https://doi.org/10.1016/B978-0-323-85387-3.00001-X>.
- [12] El Mekawy A, Hegab HM, Mohanakrishna G, Pant D, Wang H. Integrated bioelectrochemical platforms. *Biomass, Biofuels, Biochem Microb Electrochem Technol Sustain Platf Fuels, Chem Remediat* 2018;1037–58. <https://doi.org/10.1016/B978-0-444-64052-9.00043-1>.
- [13] Logan BE, Call D, Cheng S, Hamelers HVM, Sleutel THJA, Jeremiasse AW, et al. Microbial electrolysis cells for high yield hydrogen gas production from organic matter. *Environ Sci Technol* 2008;42:8630–40. <https://doi.org/10.1021/es801553z>.

- [14] Call D, Logan BE. Hydrogen production in a single chamber microbial electrolysis cell lacking a membrane. *Environ Sci Technol* 2008;42:3401–6. <https://doi.org/10.1021/es8001822>.
- [15] Lalaurette E, Thammannagowda S, Mohagheghi A, Maness PC, Logan BE. Hydrogen production from cellulose in a two-stage process combining fermentation and electrohydrogenesis. *Int J Hydrogen Energy* 2009;34:6201–10. <https://doi.org/10.1016/j.ijhydene.2009.05.112>.
- [16] Sun R, Xing D, Jia J, Liu Q, Zhou A, Bai S, et al. Optimization of high-solid waste activated sludge concentration for hydrogen production in microbial electrolysis cells and microbial community diversity analysis. *Int J Hydrogen Energy* 2014;39:19912–20. <https://doi.org/10.1016/j.ijhydene.2014.09.163>.
- [17] Pasupuleti SB, Srikanth S, Venkata Mohan S, Pant D. Development of exoelectrogenic bioanode and study on feasibility of hydrogen production using abiotic VITO-CoRETM and VITO-CASETM electrodes in a single chamber microbial electrolysis cell (MEC) at low current densities. *Bioresour Technol* 2015;195:131–8. <https://doi.org/10.1016/j.biortech.2015.06.145>.
- [18] Yossan S, Xiao L, Prasertsan P, He Z. Hydrogen production in microbial electrolysis cells: Choice of catholyte. *Int J Hydrogen Energy* 2013;38:9619–24. <https://doi.org/10.1016/j.ijhydene.2013.05.094>.
- [19] Wan LL, Li XJ, Zang GL, Wang X, Zhang YY, Zhou QX. A solar assisted microbial electrolysis cell for hydrogen production driven by a microbial fuel cell. *RSC Adv* 2015;5:82276–81. <https://doi.org/10.1039/c5ra16919d>.
- [20] Rozendal RA, Hamelers HVM, Molenkamp RJ, Buisman CJN. Performance of single chamber biocatalyzed electrolysis with different types of ion exchange membranes. *Water Res* 2007;41:1984–94. <https://doi.org/10.1016/j.watres.2007.01.019>.
- [21] Hu H, Fan Y, Liu H. Hydrogen production in single-chamber tubular microbial electrolysis cells using non-precious-metal catalysts. *Int J Hydrogen Energy* 2009;34:8535–42. <https://doi.org/10.1016/j.ijhydene.2009.08.011>.
- [22] Montpart N, Rago L, Baeza JA, Guisasola A. ScienceDirect Hydrogen production in single chamber microbial electrolysis cells with different complex substrates. *Water Res* 2014;68:601–15. <https://doi.org/10.1016/j.watres.2014.10.026>.